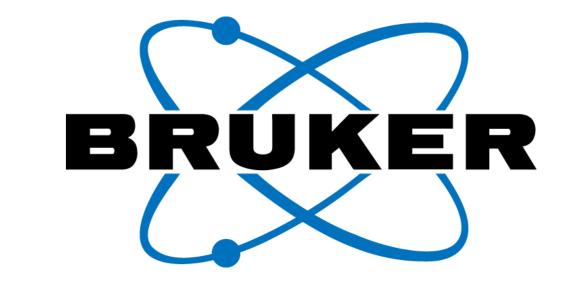
Characterization of Protein-based Biotherapeutics by TIMS enabled Next-Generation MALDI Top-Down Sequencing



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Summary

- Next-Gen MALDI-TDS: Top-down sequencing based on MALDI-ISD taking advantage of timsTOF technology
- MALDI-ISD yields 1+ N- and C-terminal backbone fragments allowing for straightforward sequence readout without charge deconvolution
- Trapped Ion Mobility Spectrometry (TIMS) separates MALDI-ISD fragments according N- or C-terminal origin enabling terminally dissected top-down sequence analysis.
- TIMS enables interference-free T³-Sequencing (CID-MS/MS of selected MALDI-ISD fragments) for verification of very terminal sequence regions.
- Next-Gen MALDI-TDS allows for rapid, in-depth characterization of proteinbased biologics re. primary sequence, terminal status and near-terminal modifications

Methods

Samples:

Nanobody: Reduction with DTT; alkylation with IAA; sample clean-up by microdialysis Recomb. SARS-CoV-2-S-glycoprotein-RBD: Expressed in and purified from HEK293 cells; recuction with DTT; release of N-glycans with PNGaseF (Promega); cleavage of sialic acids with SialExo (Genovis); sample clean-up by on-target washing

MALDI preparation:

10-20 pmol protein per sample spot; MALDI matrix sDHB (Bruker); MALDI plate MTP Anchorchip 384 BC (Bruker); red phosphorus was spotted on a separate spot position as calibrant for instrument m/z calibration Data acquisition:

Bruker timsTOF fleX instrument with dual ESI/MALDI ion source; Positive MALDI ion mode; TIMSin pressure reduced to 1.8 mbar TIMS ramp 300ms; 1/K0 range 0.6 – 1.9 Data analysis:

Bruker DataAnalysis (smoothing, baseline, peakfinding), Bruker Biotools (sequence analysis)

Results I (timsTOFfleX performance characteristics)

Performance of the timsTOF fleX instrument in Next-Gen MALDI-TDS was invstigated using Carbonic Anhydrase II (bos taurus), MW 29 kDa, as a standard protein:

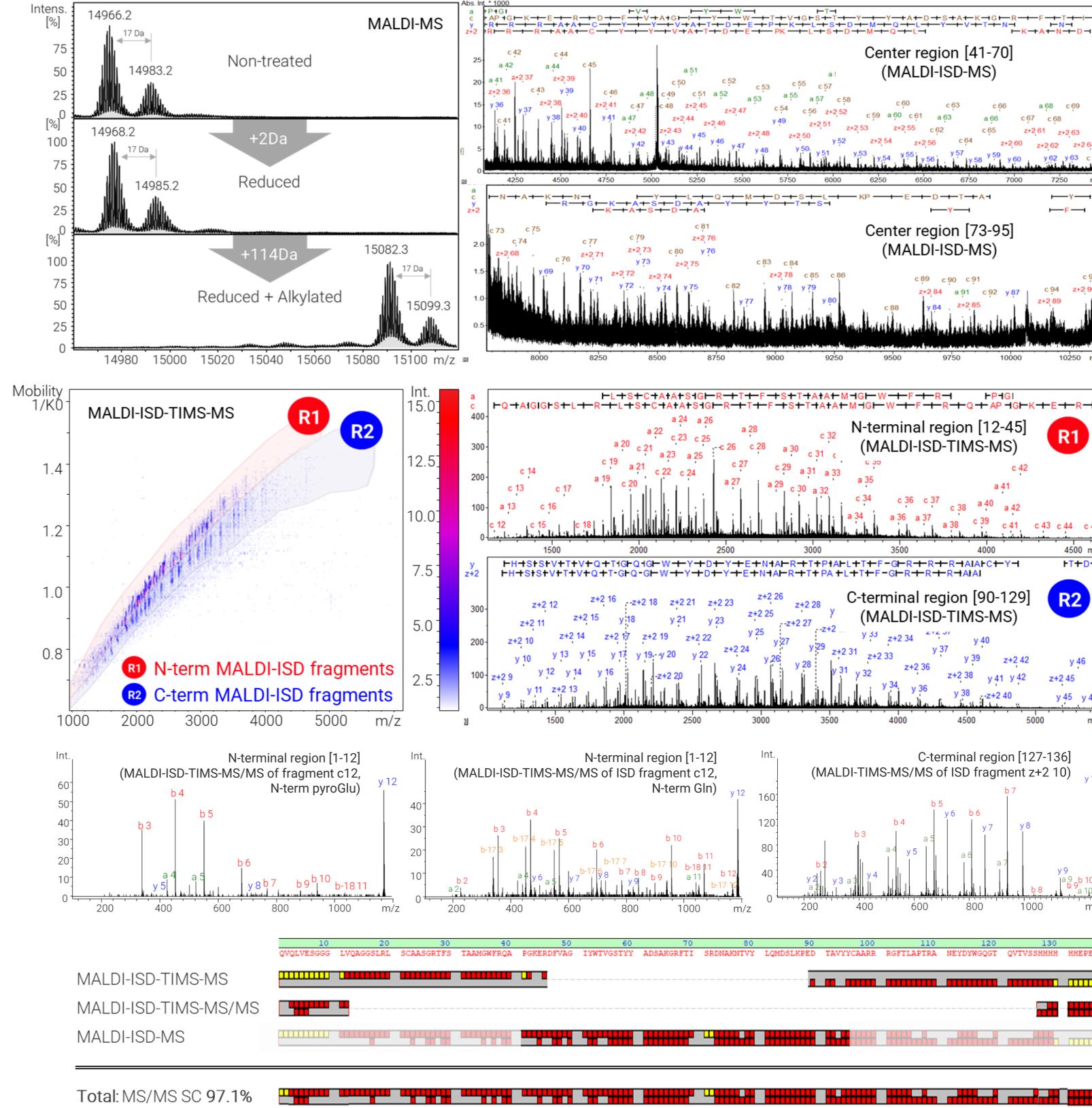
- Sequence verification percentage (SVP) achieved by MALDI-ISD-MS: 86.9%
- ISD fragments throughout m/z range 1,000 15,000 all isotopically resolved
- RMS Δm/z (across all ISD fragments): 1.67 ppm
- Acccurate intact-mass: $\Delta m/z$ -0.5 ppm ([M+3H]³⁺ signal)
- TIMS benefits to Next-Gen MALDI-TDS:
 TIMS separation of ISD fragmer
 - TIMS separation of ISD fragments according N- or C-terminal origin enables terminally dissected top-down analysis of terminal sequence regions
 - Reduced data complexity
 - Overlaps resolved for isobaric fragments originating from opposite termini
 - Simplified data interpretation in denovo-like sequencing tasks
 - Enhanced T³-Sequencing (i.e. CID-MS/MS of selected ISD fragments) of very terminal sequence regions by TIMS separation of co-isolated isobaric background

MALDI-ISD-MS $[M+3H]^{3+}$ $[M+3H]^{3+}$ (ΔMW : -0.5 ppm) [M+3H]³⁺, theoretical RMS Δm/z (ISD fragments): 11250 11350 11300 MALDI-ISD-TIMS--MS C-term ISD fragments (1+) N-term ISD fragments 1+ T³-Sequencing (MALDI-ISD-TIMS-CID-MS/MS) 200 SHHWGYGKHN GPEHWHKD 2250 Mobility 1/K0 1.200 1.150 1.100

Results II (Biologics applications)

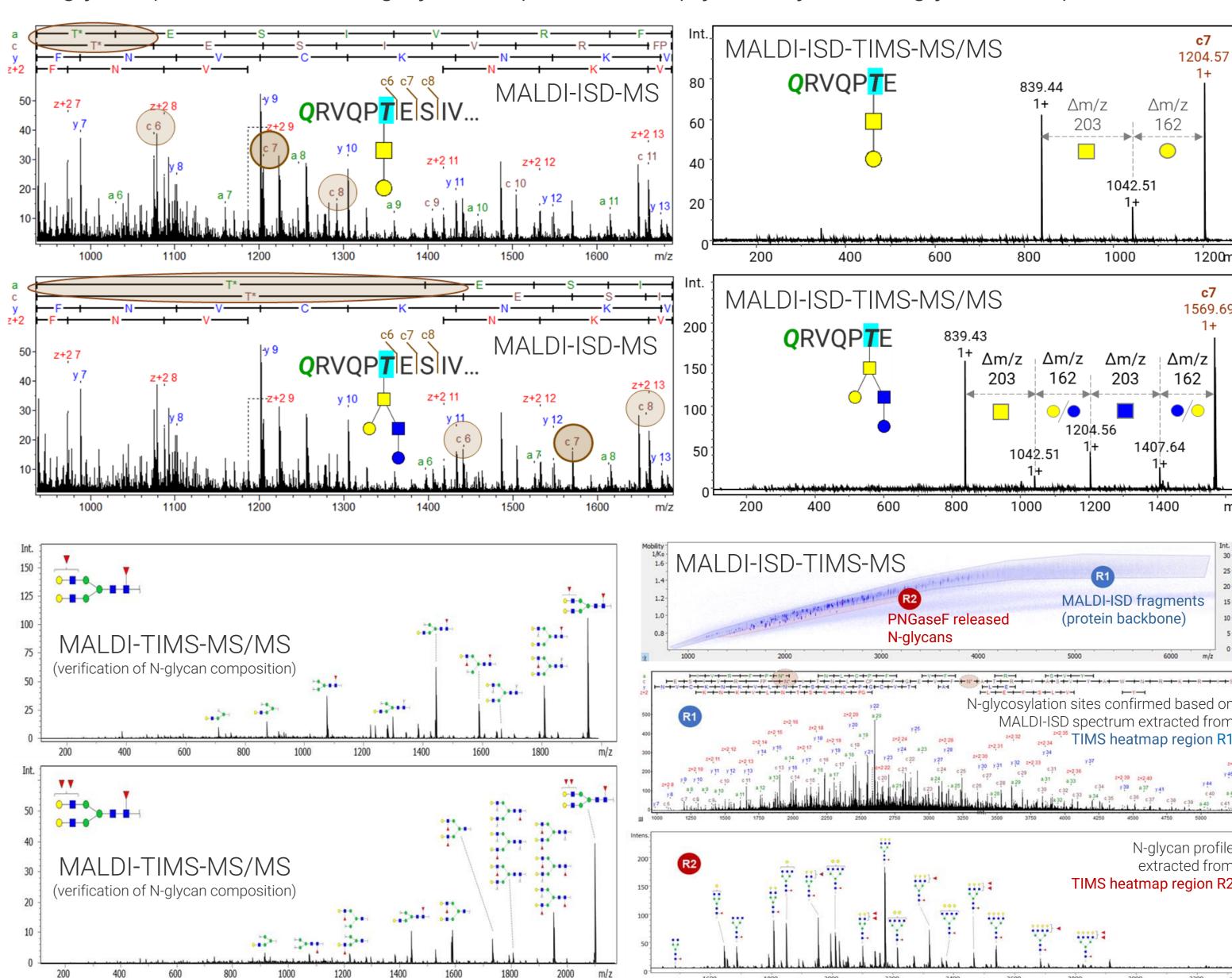
Application I: Complete characterization of a 15 kDa nanobody

- Next-Gen MALDI-TDS yielded the complete nanobody sequence
 - 136 AA residues
 - Partially Gln->pyroGlu converted N-terminus
 - 2 cysteines linked via disulfide bridge
- Overall MS/MS Seq.Cov. > 97% (132 out of 136 AA residues assigned by fragment ions)
- 20% of the AA sequence was retrieved denovo (due to conflicting/lacking reference data)
- Sequencing result reconfirmed by matching nanobody MW obtained from intact-mass MALDI-MS data (ΔMW < 0.2 ppm)



Application II: Characterization of O- and N-glycosylation in recomb. SARS-CoV-2-S-glycoprotein-RBD expressed in HEK293

- Active site of O-glycosylation pinpointed at position T6 (S8 is not O-glycosylated)
- O-glycosylation site T6 features both, core 1 and core 2 O-glycans
- Active sites of N-glycosylation verified (N14; N26)
- N-glycan profile features highly host-specific multiply fucosylated N-glycan compositions



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Biologics / timsTOF fleX